Cite this article: Abbas, Z. I., Sabbar, A. G., & Obaid, W. K. (2025). Correlation between serum levels of cyclooxygenase II and microsomal prostaglandin E synthase 1 in Iraqi colorectal cancer patients and healthy controls. *Journal of Current Science and Technology*, 15(4), Article 142. https://doi.org/10.59796/jcst.V15N4.2025.142



Journal of Current Science and Technology

Journal homepage: https://jcst.rsu.ac.th



Correlation between Serum Levels of Cyclooxygenase II and Microsomal Prostaglandin E Synthase 1 in Iraqi Colorectal Cancer Patients and Healthy Controls

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Received 1 June 2025; Revised 23 June 2025; Accepted 2 July 2025; Published online 20 September 2025

Abstract

Colorectal cancer (CRC) ranks among the most widespread cancers worldwide, exhibiting considerable mortality rates. Chronic inflammation plays a crucial role in driving tumor development. Cyclooxygenase-II is an inducible enzyme that aids in the production of prostaglandins using arachidonic acid, particularly during inflammatory responses. Microsomal prostaglandin E synthase-1, functioning downstream of COX-2, specifically converts PGH2 to PGE2, and correlates with unfavorable outcomes in CRC. While their roles have been well documented in tissue, limited research has assessed their levels in serum. The present study aimed to evaluate the serum levels of COX-2 and mPGES-1 in CRC patients compared to healthy controls, and to investigate the correlation between these biomarkers across all participants, regardless of clinical grouping. A total of 70 patients, 35 newly diagnosed and 35 undergoing treatments were included, along with 30 healthy individuals as controls. Blood samples were collected from healthy individuals and Iraqi CRC patients. Serum concentrations of COX-2 and mPGES-1 were measured using ELISA. Marked variations in the levels of these enzymes were noted among the examined groups, with newly diagnosed patients showing the highest levels compared to treated patients and healthy control. A robust positive correlation was observed between levels of COX-2 and mPGES-1 in all groups. These outcomes suggest that serum levels of Cyclooxygenase II and Microsomal Prostaglandin E synthase 1 could serve as prospective diagnostic indicators for CRC patients.

Keywords: serum biomarkers; COX-2; mPGES-1; CRC, ELISA.

1. Introduction

Colorectal cancer (CRC) poses a critical public health issue, noted for its high incidence and mortality rates worldwide. CRC is the third most prevalent cancer and the second leading cause of cancer deaths. Around 1.8 million new diagnoses and 881,000 fatalities were recorded in 2018 (Baidoun et al., 2021). In Iraq, the overall incidence rate rose from 2.28 to 6.18 per 100,000 individuals from 2000 to 2019, with a notable increase among younger patients aged 20 to 50 years (Alwan et al., 2022). Similarly, mortality

rates increased, from 1.25 to 1.77 per 100,000 during the same period (Ibrahem et al., 2022). Long-term inflammation is recognized as a major contributor to the onset and advancement of various cancers, involving colorectal carcinoma (Grivennikov et al., 2010; Masae et al., 2025). A primary pathway connecting inflammation to cancer is arachidonic acid (AA). This fatty acid originates from membrane phospholipids and is metabolized by cyclooxygenases (COXs) and other enzymes into various bioactive lipids, such as PGE2 and leukotrienes, which are

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associated with CRC progression (Ricciotti & FitzGerald, 2011; Mohammed & Shoemaker, 2022; Gretschel et al., 2024). Therefore, the interaction between chronic inflammation and AA metabolism environment favorable fosters an progression. Cyclooxygenase (COX) necessary steps in the production of prostanoids (Ekhlas et al., 2022). In particular, COX-2 is an inducible enzyme that facilitates the conversion of arachidonic acid into a volatile prostaglandin H₂ (Nanda & Dhawan, 2021). Previous research found that CRC tissues displayed significantly higher levels of COX-2 mRNA compared to adjacent normal tissues (Negi et al., 2019). Moreover, this enzyme was found to influence tumor initiation, progression, metastasis, and prognosis (Albasri et al., 2018). Other isoforms of cyclooxygenase include COX-1, represents the constitutive variant, and COX-3, which is mainly found in the brain and spinal cord (Fitzpatrick, 2004; Davies et al., 2004).

Microsomal prostaglandin E synthase 1 (PGES 1) is an enzyme that can be triggered by inflammatory response, facilitating the transformation prostaglandin H2 into PGE2 into several cancers (Korotkova & Jakobsson, 2014; Kamei et al., 2010). The other isoforms of PGES include microsomal PGES-2 and cytosolic PGES (cPGES) (Gudis et al., 2005). Unlike mPGES-1, mPGES-2 is constitutively expressed across various tissues and shows minimal regulation from inflammatory signals (Murakami & Kudo, 2006). In colorectal cancer, mPGES-1 is frequently co-expressed with COX-2, and is increasingly recognized as a critical driver of CRC development and progression (Sasaki et al., 2015; Stamatakis et al., 2015; Sabah & Hassan, 2024). Furthermore, the heightened expression of PGES, especially the microsomal variant, has been observed in colorectal adenomas and cancers, resulting in increased PGE2 levels (Kim & Kim, 2021). The high levels of PGE2 can promote the development and invasion of tumor through many mechanisms, including stimulating cell proliferation, immunosuppression, preventing apoptosis, regulating angiogenesis, enhancing cell migration, and contributing to drug resistance (Karpisheh et al., 2019; Sheng et al., 2020). COX-2 and mPGES-1 represent promising targets for diagnosis, prognosis, and therapeutic strategies in colorectal Overexpression of COX-2 in CRC patients has been associated with to lymph node and liver metastasis, along with poor prognosis (Purnama et al., 2023). Moreover, previous studies showed that genetic deletion or suppression of mPGES-1 in mice reduced tumor growth by approximately 80-90% (Yarla et al., 2020). Furthermore, mPGES-1 inhibitors may offer a promising target to avoid cardiovascular side effects that are associated with COX-2 inhibitors (Yarla et al., 2020). Previous studies have focused on tissue-based methods such as immunohistochemistry (IHC) or polymerase chain reaction (PCR), while serum-based studies remain limited, particularly in Iraqi populations.

2. Objective

This study intends to measure the serum levels of Cyclooxygenase II (COX-2) and microsomal prostaglandin E synthase 1 (mPGES-1) in CRC patients compared with healthy controls using enzyme-linked immunosorbent assay (ELISA). It also explores the relationship between these two biomarkers.

3. Materials and Methods

3.1 Study Design

This case-control study consisted of 70 CRC patients and 30 apparently healthy individuals, with blood samples and data collected from Al-Amal Hospital for Radiation and Nuclear Medicine and the Digestive and Liver Disease Teaching Hospital in Medical City, Baghdad, Iraq, from January until April 2025. The studied groups were primarily categorized into 3 groups: Group I: included 35 newly diagnosed patients (without treatment, covering all stages). Group II: included 35 CRC patients undergoing medication (chemotherapy and radiotherapy). Group III: consisted of 30 healthy volunteers with no history of cancer or any gastrointestinal problems such as inflammatory bowel disease (IBD), celiac disease, or GI infections. Patients were carefully selected based on specific inclusion and exclusion standards. The inclusion criteria included newly diagnosed CRC patients and patients who had received treatment (chemotherapy and radiotherapy). Exclusion criteria included individuals with conditions that could influence the inflammatory biomarkers, such as diabetes, heart disease, kidney failure, and gastrointestinal disorders.

A comprehensive set of parameters, including age, gender, smoking status, and treatment regimen, was collected from patients' medical records. The necessary permissions were obtained from Al-Amal Hospital for Radiation and Nuclear Medicine and the Digestive and Liver Disease Teaching Hospital in Medical City. All patients in the medication group received at least two cycles of chemotherapy. Treatment regimens included standard chemotherapy

combinations (e.g., FOLFOX, XELOX) and/ or radiotherapy, with some patients receiving biological agents such as AVASTIN. In this study, the sample size was determined utilizing G*Power software version 3.1. An effect size of f = 0.5 was assumed due to the absence of consistent local data. This large effect size assumption was supported by previous literature (Han et al., 2014; Yang et al., 2018). The analysis indicated that a minimum of 18 participants per group was sufficient to provide meaningful results. However, a total sample size of 100 participants was chosen based on practical considerations and potential variation. Ethical approval was obtained by the Institutional Review Board (IRB) of Middle Technical University, Iraq (approval number: 71; date:11/01/2025, P. C. No.10074). The study was conducted with full respect for participants' rights, in line with the ethical authorization of the Declaration of Helsinki. All participants provided written consent before any sample was taken.

3.2 Estimation of Serum COX-2 and mPGES-1 Levels

Blood samples were obtained from Iraqi patients with colorectal cancer and healthy individuals; 10 mL of blood was drawn intravenously, transferred to a vacuum gel tube, and allowed to stand for approximately 15 to 20 minutes to clot. Afterward, serum samples were obtained through centrifugation for 15 minutes at 1000×g. After being divided into smaller portions, all sera were frozen at -80°C until ready for use. The concentrations of the studied enzymes were measured using commercial ELISA® kits: Human COX-2 ELISA kit (ELK Biotechnology, USA; Cat. No. ELK2096) and human mPGES-1 ELISA kit (Cloud-Clone Corp, USA; Cat. No. SEA167Hu). Absorbance was measured using a human reader HS microplate reader (human GmbH, Germany). Both assays utilized a sandwich enzyme immunoassay method. For COX-2, 100 µL of each serum sample or standard was placed in the wells and incubated at 37°C for 80 minutes, followed by incubating with a biotin-conjugated detection antibody for 50 minutes, washing, and then adding streptavidin-HRP for another 50 minutes. After the final washes, the TMB substrate was added and kept in the dark. The reaction was stopped with sulfuric acid, and optical density (OD) was measured at 450 nm using a microplate reader. For mPGES-1, a similar procedure was followed with minor variations in incubation times. Samples were incubated for 2 hours at 37°C. All measurements were performed in

duplicate, and concentrations were determined using a standard curve.

3.3 Statistical Analysis

SPSS (2019) program was used to assess the impact of various sets (patients and control group) on the study variables. An independent t-test and one-way ANOVA with LSD post-hoc analysis were conducted for COX-2 and mPGES-1 levels after confirming the normality and homogeneity of variances. The Chi-square test was used to evaluate significant differences in percentages at probability levels of 0.05 and 0.01. The correlation coefficient was calculated to examine the relationship between COX-2 and mPGES-1.

4. Results and Discussion

4.1 The Demographic Characteristics of the Studied Groups

Seventy patients were divided into two groups according to their treatment status: 35 CRC newly diagnosed patients with colorectal cancer and 35 patients diagnosed with CRC and undergoing treatment. The control group consisted of 30 apparently healthy participants.

In this study males represented a slightly higher proportion (54.29%) compared to females (45.71%). However, the differences in sex distribution were not statistically significant among the studied groups (p = 0.502, NS). These findings are consistent with several Iraqi studies (Al-Saigh et al., 2019; Radhi et al., 2018, Falih et al., 2020), which reported no significant sex differences among CRC patients. A previous literature suggested that although sex may influence CRC risk factors, it may not always be reflected in the distribution across clinical groups (Siegel et al., 2023).

Individuals under 50 years represent the young-onset CRC group, a subgroup that has shown a rising incidence globally (Siegel et al., 2020). The 50-60 years age group represents a transitional group, aligning with the age threshold for initiating CRC screening (Shaukat et al., 2021). Meanwhile, individuals above 60 years represent the population with the highest incidence of CRC. The distribution of study participants across age groups is summarized in Table 1. Most newly diagnosed CRC patients (60%) were above 60 years old. In contrast, patients undergoing treatment were mostly between 50-60 years old (37. 14%). The control group had a large proportion of younger participants (43.33%). Moreover, the younger age group (<50 years) had fewer new diagnoses (8.57%) compared to other age groups. The results revealed

a significant association between age groups and CRC treatment status (p = 0.0267) were significantly associated. The mean age for each group showed that the newly diagnosed patients were significantly older (62.11 \pm 2.08) than both patients undergoing treatment and control patients (53.91 \pm 2.32, 52.17 \pm 2.83) respectively. These findings are consistent with both the annual report of Iraqi Cancer Registry (2023) and a registry-based study conducted over two decades in Iraq, confirming that CRC incidence in Iraq is lowest under 50, intermediate in 50-60 bracket, and highest above 60 (Ibrahem et al., 2022). The control group had the youngest mean age, as it reflects the healthy population. The noted age differences among the studied groups- particularly in newly diagnosed patients' group - may influence the inflammatory pathways, which are central to colorectal cancer development, as aging is associated with a chronic, low-grade inflammation known as "inflammaging"

(Serrano-López & Martín-Antonio, 2021). Therefore, age may act as a confounding factor by partially contributing to the elevation of inflammatory biomarkers.

The association between cigarette smoking and colorectal cancer (CRC) was non-significant among the studied groups (p = 0.086, NS). As presented in Table 1, these findings mirror the results of a previous descriptive study conducted in Iraq (Al-Saigh et al., 2019). The association between CRC and smoking may vary based on tumor characteristics, such as immune response, indicating that the relationship is not uniform across all cases (Hamada et al., 2019). Moreover, meta-analysis research indicated that the risk was greater in nations with higher Human Development Index (HDI), suggesting that socioeconomic factors may influence this relationship (Keivanlou et al., 2023).

Table 1 Groups of patients with CRC categorized by age and gender, with the group of healthy persons as the control

Factor		New diagnosis No. (%)	Undergoing treatment No. (%)	Control No. (%)	p-value
Gender	Male	19 (54.29%)	19 (54.29%)	16 (53.33%)	0.502 ^{NS}
Genuer	Female	16 (45.71%)	16 (45.71%)	14 (46.67%)	
	<50 yr.	3 (8.57%)	11 (31.43%)	13 (43.33%)	
Age groups	50-60 yr.	11 (31.43%)	13 (37.14%)	9 (30.00%)	
(year)	>60 yr.	21 (60.00%)	11 (31.43%)	8 (26.67%)	0.0267 *
	$Mean \pm SE$	62.11 ± 2.08 a	53.91 ± 2.32 b	52.17 ± 2.83 b	0.0087 **
Cmakina	Yes	12 (34.29%)	9 (25.71%)	8 (26.67%)	$0.086~^{ m NS}$
Smoking	No	23 (65.71%)	26 (74.29%)	22 (73.33%)	
To	Total		35	30	

Values are presented as number (%) and Mean \pm SE.

Means in the same column followed by different superscript letters differ significantly. * $p \le 0.05$; ** $p \le 0.01$; NS = not significant.

Table 2 Mean serum levels of COX-2 and mPGES-1 in different study groups

C	$Mean \pm SE$		
Group —	COX-2	mPGES-1	
Newly diagnosed	4.49 ± 0.06 a	53.74 ± 1.79 a	
Inder medication	3.27 ± 0.03 b	28.90 ± 1.23 b	
Control	2.07 ± 0.04 °	19.33 ± 0.32 °	
L.S.D.	0.139 **	3.736 **	
p-value	0.0001	0.0001	
	** (p≤0.01)		

Values are presented as Mean \pm SE.

Means in the same column followed by different superscript letters differ significantly (** $p \le 0.01$).

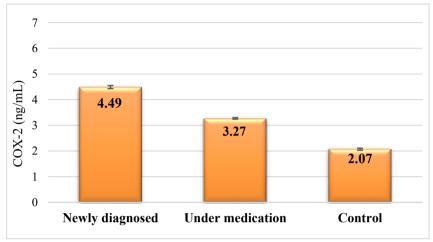


Figure 1 Comparison of serum COX-2 levels (ng/mL) among newly diagnosed CRC patients, patients under medication, and healthy controls

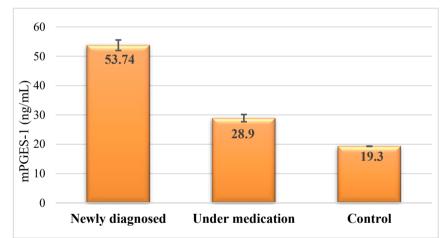


Figure 2 Comparison of serum mPGES-1 levels (ng/mL) among newly diagnosed CRC patients, patients under medication, and healthy controls

4.2 Evaluation of Cyclooxygenase II

Cyclooxygenase II (COX-2) is an essential enzyme in the arachidonic acid pathway, converting arachidonic acid into prostaglandins and thromboxanes. COX-2 mainly functions in inflammatory reactions; however, it also produces pro-resolving metabolites that contribute to the resolving inflammation (Chun et al., 2024; Faki & Er, 2021). In this study, the serum levels of COX-2 were significantly higher in CRC patients compared with healthy controls. The mean COX-2 concentration in newly diagnosed CRC patients was 4.49 ± 0.06 , while in patients undergoing treatment, it was 3.27 ± 0.03 . In contrast, the mean COX-2 level in the control group was 2.07 ± 0.04 . The results revealed highly significant differences among the three groups (p = 0.0001). Post-hoc comparisons

confirmed significant differences between all groups. Although the differences in COX-2 levels may appear slight potentially due to variations in ELISA kits used, including differences in detection range, sensitivity, and calibration standards they were statistically significant between CRC patient groups and healthy controls (p = 0.0001). Comparable findings were reported in a recent Iraqi study (Mowat & Al-Abady, 2025). Additionally, other ELISA-based studies (Cao et al., 2019; Yang et al., 2018; Choosang et al., 2024) were also consistent with these findings. However, as shown in Figure 1, the newly diagnosed patients in this study exhibited higher mean COX-2 level when compared with CRC patients undergoing treatment. This increase may result from COX-2 overexpression in cancer cells, which is associated with tumor

severity and poor clinical outcomes (Szweda et al., 2019). In the same context, a recent study by Shalaby et al., (2025) found that chemo-resistant colon cancer tissues exhibited a significant increase in COX-2 mRNA compared with tissues that did not undergo chemotherapy (p < 0.001). In contrast, this study showed a significant decrease in serum COX-2 levels in post-treatment patients. This is consistent with observations that responding tumors exhibit lower COX-2 levels than resistant cases (Rahman et al., 2012; Sophonnithiprasert et al., 2025). While this study used serum ELISA to quantify COX-2, previous studies (Negi et al., 2019; Roelofs et al., 2014) based on immunohistochemistry (IHC) and real-time quantitative polymerase chain reaction (qPCR) reported elevated COX-2 levels in CRC tissues compared with adjacent normal tissue, suggesting a potential link between tissue and circulating concentrations. While this study used serum ELISA to quantify COX-2, previous studies (Negi et al., 2019; Roelofs et al., 2014) based on immunohistochemistry (IHC) and real-time quantitative polymerase chain reaction (qPCR) reported elevated COX-2 levels in CRC tissues compared with adjacent normal tissue, suggesting a potential link between tissue and circulating concentrations. COX-2 expression in healthy colon tissues is typically lower than in CRC tissues. Nevertheless, several studies have reported that COX-2 expression is still detectable in healthy colon tissues, especially in patients with a family history of CRC or other risk factors (Lin et al., 2013).

4.3 Evaluation of mPGES-1

Microsomal prostaglandin E synthase 1 (mPGES-1) is an inducible enzyme that plays a crucial role in the production of PGE2 under inflammatory conditions (Kojima et al., 2024). As shown in Table 2 and Figure 2, mPGES 1 exhibited highly significant differences among the groups examined (p = 0.0001). CRC patients without treatment exhibit higher mPGES-1 levels compared with patients undergoing treatment (p = \leq 0.001). Although ELISA-

based studies on quantifying mPGES-1 in CRC patients are limited, a parallel study employing realtime polymerase chain reaction (RT-PCR) to measure PTGES expression reported that PTGES levels were markedly increased in CRC cells compared with normal cells (Geng et al., 2023). This finding is in agreement with our results. Moreover, mPGES-1 expression has been linked to the development of polyps that emerge during the early stage of colon cancer (Myung et al., 2006). Subsequent studies have found that mPGES-1 overexpression is associated with significantly poorer prognosis in CRC patients from stages I to III (Seo et al., 2009). The elevated levels of mPGES-1 in CRC patients may result from its strong association with COX-2, which is frequently overexpressed in CRC, thereby leading to higher PGE₂ levels (Stamatakis et al., 2015). Inflammatory signals, such as TNF- α and other cytokines within the tumor microenvironment, further enhance mPGES-1 expression. thereby contributing progression (Kim & Kim, 2021). within the tumor microenvironment, further enhance mPGES-1 expression, thereby higher in both serum and tissue samples of CRC patients than in healthy controls. Furthermore, the present study observed a significant elevation in serum mPGES-1 levels, which mainly participate in PGE₂ synthesis. This evidence further supports the potential utility of mPGES-1 as a noninvasive biomarker for detecting or monitoring CRC. Additionally, the results of this study also indicate that CRC patients who had received chemotherapy and/or radiotherapy exhibited lower blood levels of mPGES-1 compared with untreated individuals. Although the comparison involved two different patient groups rather than longitudinal samples, the distinction indicates that anti-cancer treatment may be associated with reduced mPGES-1 levels.

4.4 Cyclooxygenase II and mPGES-1 Association

Spearman's correlation analysis was performed to determine the potential relationship between Cyclooxygenase II (COX-2) and mPGES-1.

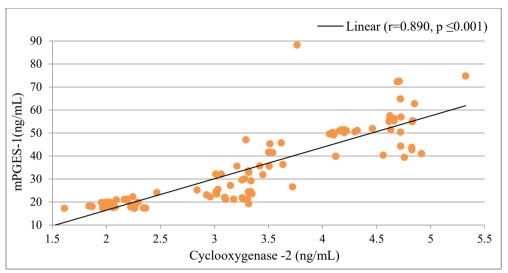


Figure 3 Linear correlation between serum COX-2 and mPGES-1 levels (ng/mL) across all study groups $(r = 0.890, p \le 0.001)$

Figure 3 illustrates the correlation between those two biomarkers across all study groups. A very strong and highly statistically significant positive correlation was observed (r = 0.890, $p = \le 0.001$). This association was consistent across newly diagnosed patients, patients undergoing treatment, and healthy individuals.

The findings revealed a significant positive correlation between COX-2 and microsomal Prostaglandin E synthase-1 (mPGES-1) in the studied groups, mainly due to COX-2's function in facilitating tumorigenesis via prostaglandin-mediated signaling pathways.

COX-2 is a crucial enzyme that facilitates the conversion of arachidonic acid (AA) prostaglandin H₂ (PGH₂), which is subsequently transformed into PGE2 by mPGES-1. In colorectal cancer, elevated COX-2 expression is commonly observed and is associated with tumor severity and poor clinical outcomes (Finetti et al., 2023), mPGES-1 acts as a final synthase that works downstream of COX-2 to produce PGE₂ (Hyodo et al., 2022). It is highly expressed in CRC and functionally associated with COX-2, particularly under inflammatory conditions (Stamatakis et al., 2015). The regulation of COX-2 and mPGES-1 occurs in a coordinated manner in colorectal cancer. The upregulation of COX-2 leads to increased mPGES-1 expression through an early growth response 1 (EGR1)-dependent mechanism, which is further facilitated by prostaglandins such as PGF₂α produced downstream of COX-2 (Stamatakis et al., 2015). A previous study by Seo et al., (2009)

analyzed prostaglandin E synthase (PGES) isomers and COX-2 in CRC tissues using different sample types and detection methods (RT-qPCR, Western blot, and IHC). They reported that mRNA and protein levels of mPGES-1 were overexpressed in CRC tissues compared with paired normal tissues and were correlated with poorer prognosis in stage I-III CRC patients. While their study focused on tissue samples, the present study detected higher serum levels of COX-2 and mPGES-1 in serum using ELISA. Despite the differences in sample types and detection methods, the current study supports their findings, demonstrating significantly higher serum levels of both biomarkers in CRC patients. Additionally, this study observed a strong positive correlation between COX-2 and mPGES-1 (r = 0.89, p ≤ 0.001 ; Figure 3), suggesting a coordinated regulation of these enzymes in the systemic circulation of CRC patients. Furthermore, these biomarkers could serve as potential tools for personalized treatment planning. A potentially effective therapeutic strategy may involve targeting the COX-2/mPGES-1 pathway. Although COX-2 inhibitors are effective, they have been associated with cardiovascular and gastrointestinal side effects. Selective mPGES-1 inhibitors are currently being developed as safer therapeutic alternatives (Muthukumaradoss et al., 2022).

5. Conclusion

It can be concluded that serum concentrations of COX-2 and mPGES-1 varied significantly among all participants. Interestingly, the group of newly

diagnosed CRC patients exhibiting the highest levels compared with both undergoing treatment patients and healthy controls. The strong positive correlation between these two biomarkers further highlights their interconnected roles in CRC-associated inflammation and progression. Overall, these data showed that COX-2 and mPGES-1 may serve as potential diagnostic biomarkers for colorectal cancer. Further longitudinal studies with larger cohorts, treatment-specific analyses, and disease-stage stratification are recommended to validate their clinical utility.

6. Acknowledgements

We are sincerely grateful to the staff, healthcare professionals, and administration of the Al-Amal Hospital for Radiation and Nuclear Medicine and the Digestive and Liver Disease Teaching Hospital at Baghdad Medical City for their unwavering dedication, exceptional patient care, and invaluable assistance in providing and collecting the samples required for this research.

7. CRediT Statement

Zainab I. Abbas: Conceptualization, methodology, data curation, writing, original draft, visualization, resources.

Ahmed Gh. Sabbar: Supervision, formal analysis, investigation, writing, review & editing.

Wajeeh K. Obaid: Supervision, writing, review & editing, project administration, validation.

8. References

- Albasri, A. M., Elkablawy, M. A., Hussainy, A. S., Yousif, H. M., & Alhujaily, A. S. (2018). Impact of cyclooxygenase-2 over-expression on the prognosis of colorectal cancer patients: An experience from Western Saudi Arabia. *Saudi Medical Journal*, *39*(8), 773-780. https://doi.org/10.15537/smj.2018.8.22837
- Al-Saigh, T., Al-Bayati, S., Abdulmawjood, S., & Ahmed, F. (2019). Descriptive study of colorectal cancer in Iraq, 1999-2016. *Annals of the College of Medicine Mosul*, 41(1), 81–85.
- Alwan, N. A., Lami, F., Al Nsoor, M., & Kerr, D. (2022). Trends in the incidence and mortality of the most common cancers in Iraq (Iraqi cancer registry 1999-2019). *The Gulf Journal of Oncology*, *1*(40), 47-57.
- Baidoun, F., Elshiwy, K., Elkeraie, Y., Merjaneh, Z., Khoudari, G., Sarmini, M. T., ... & Saad, A. (2021). Colorectal cancer epidemiology: recent trends and impact on outcomes.

- Current Drug Targets, 22(9), 998-1009. https://doi.org/10.2174/1389450121999201117115717
- Cao, H., Wang, Q., Gao, Z., Xu, X., Lu, Q., & Wu, Y. (2019). Clinical value of detecting IQGAP3, B7-H4 and cyclooxygenase-2 in the diagnosis and prognostic evaluation of colorectal cancer. *Cancer Cell International*, 19(1), Article 163. https://doi.org/10.1186/s12935-019-0881-3
- Choosang, K., Boonsilp, S., Kritsiriwuthinan, K., Chumchuang, P., Thanacharoensakun, N., Saai, A., & Pongparit, S. (2024). A dot-blot ELISA preliminary evaluation using PvMSP1-42 recombinant protein as antigen for serological diagnosis of Plasmodium vivax infection in Thailand. *European Journal of Microbiology and Immunology*, *14*(2), 202-209. https://doi.org/10.1556/1886.2024.00008
- Chun, K. S., Kim, E. H., Kim, D. H., Song, N. Y., Kim, W., Na, H. K., & Surh, Y. J. (2024). Targeting cyclooxygenase-2 for chemoprevention of inflammation-associated intestinal carcinogenesis: An update. *Biochemical Pharmacology*, 228, Article 116259.
- https://doi.org/10.1016/j.bcp.2024.116259
 Davies, N. M., Good, R. L., Roupe, K. A., & Yáñez, J. A. (2004). Cyclooxygenase-3: Axiom, dogma, anomaly, enigma or splice error? Not as easy as 1, 2, 3. *Journal of Pharmacy & Pharmaceutical Sciences*, 7(2), 217-226.
- Ekhlas, S. F., Hameed, S. H. O., & Hameed, F. R. (2022). Evaluation of circulating microRNA-221 and IL-10 levels as potential diagnostic markers for colorectal cancer in correlation with prognosis. *International Journal of Drug Delivery Technology*, 12(3), 129–137. https://doi.org/10.25258/ijddt.12.3.23
- Faki, Y., & Er, A. (2021). Different chemical structures and physiological/pathological roles of cyclooxygenases. *Rambam Maimonides Medical Journal*, *12*(1), Article e0003. https://doi.org/10.5041/RMMJ.10426
- Falih, E. S., Obaid, S. H., & Hameed, F. R. (2020). Evaluation of the role of miRNA-21 levels as a potential diagnostic biomarker for colorectal cancer associated with prognosis. *Medicolegal Update*, 20(2), Article 509.
- Finetti, F., Paradisi, L., Bernardi, C., Pannini, M., & Trabalzini, L. (2023). Cooperation between prostaglandin E2 and epidermal growth factor

- receptor in cancer progression: A dual target for cancer therapy. *Cancers*, *15*(8), Article 2374. https://doi.org/10.3390/cancers15082374
- Fitzpatrick, F. A. (2004). Cyclooxygenase enzymes: regulation and function. *Current Pharmaceutical Design*, *10*(6), 577-588. https://doi.org/10.2174/1381612043453144
- Geng, S., Zhan, H., Cao, L., Geng, L., & Ren, X. (2023). Targeting PTGES/PGE2 axis enhances sensitivity of colorectal cancer cells to 5-fluorouracil. *Biochemistry and Cell Biology*, 101(6), 501-512. https://doi.org/10.1139/bcb-2023-0101
- Gretschel, J., El Hage, R., Wang, R., Chen, Y., Pietzner, A., Loew, A., ... & Schmöcker, C. (2024). Harnessing oxylipins and inflammation modulation for prevention and treatment of colorectal cancer. *International Journal of Molecular Sciences*, 25(10), Article 5408. https://doi.org/10.3390/ijms25105408
- Grivennikov, S. I., Greten, F. R., & Karin, M. (2010). Immunity, inflammation, and cancer. *Cell*, *140*(6), 883-899. https://doi.org/10.1016/j.cell.2010.01.025
- Gudis, K., Tatsuguchi, A., Wada, K., Futagami, S., Nagata, K., Hiratsuka, T., ... & Sakamoto, C. (2005). Microsomal prostaglandin E synthase (mPGES)-1, mPGES-2 and cytosolic PGES expression in human gastritis and gastric ulcer tissue. *Laboratory Investigation*, 85(2), 225-236. https://doi.org/10.1038/labinvest.3700200
- Han, X., Li, H., Su, L., Zhu, W., Xu, W., Li, K., ... & Liu, H. (2014). Effect of celecoxib plus standard chemotherapy on serum levels of vascular endothelial growth factor and cyclooxygenase-2 in patients with gastric cancer. *Biomedical Reports*, 2(2), 183-187. https://doi.org/10.3892/br.2013.209
- Hamada, T., Nowak, J. A., Masugi, Y., Drew, D. A., Song, M., Cao, Y.,... & Ogino, S. (2019). Smoking and risk of colorectal cancer subclassified by tumor-infiltrating T cells. *Journal of the National Cancer Institute*, 111(1), 42–51. https://doi.org/10.1093/jnci/djy059
- Hyodo, T., Ito, Y., Hosono, K., Uematsu, S., Akira, S., Majima, M., Takeda, A., & Amano, H. (2022). The role of mPGES-1 in promoting granulation tissue angiogenesis through regulatory T-cell accumulation. *In Vivo*, *36*(5), 2061–2073. https://doi.org/10.21873/invivo.12932

- Ibrahem, S., Ahmed, H., & Zangana, S. (2022).

 Trends in colorectal cancer in Iraq over two decades: Incidence, mortality, topography and morphology. *Annals of Saudi Medicine*, 42(4), 252-261. https://doi.org/10.5144/0256-4947.2022.252
- Iraqi Cancer Registry. (2023). *Ministry of Health* (*Iraq*). Global Health Data Exchange. https://ghdx.healthdata.org/organizations/ministry-health-iraq
- Kamei, D., Murakami, M., Sasaki, Y., Nakatani, Y., Majima, M., Ishikawa, Y., ... & Kudo, I. (2010). Microsomal prostaglandin E synthase-1 in both cancer cells and hosts contributes to tumour growth, invasion and metastasis. *Biochemical Journal*, 425(2), 361-371. https://doi.org/10.1042/BJ20090045
- Karpisheh, V., Nikkhoo, A., Hojjat-Farsangi, M., Namdar, A., Azizi, G., Ghalamfarsa, G., ... & Jadidi-Niaragh, F. (2019). Prostaglandin E2 as a potent therapeutic target for treatment of colon cancer. *Prostaglandins & Other Lipid Mediators*, 144, Article 106338. https://doi.org/10.1016/j.prostaglandins.2019. 106338
- Keivanlou, M. H., Amini-Salehi, E., Hassanipour, S., Mahapatro, A., Raghuma, N., Joukar, F., ... & Mansour-Ghanaei, F. (2023). Association between smoking and colorectal cancer in Eastern Mediterranean Regional Office (EMRO): A systematic review and meta-analysis. *Saudi Journal of Gastroenterology*, 29(4), 204-211. https://doi.org/10.4103/sjg.sjg 163 23
- Kim, Y. H., & Kim, K. J. (2021). Upregulation of prostaglandin E2 by inducible microsomal prostaglandin E synthase-1 in colon cancer. *Annals of Coloproctology*, *38*(2), 153-159. https://doi.org/10.3393/ac.2021.00374.0053
- Kojima, F., Hioki, Y., Sekiya, H., Kashiwagi, H., Iizuka, Y., Eto, K., ... & Ichikawa, T. (2024). Microsomal prostaglandin E synthase-1 controls colonic prostaglandin E2 production and exerts a protective effect on colitis induced by trinitrobenzene sulfonic acid in mice. *International Journal of Molecular Sciences*, 25(22), Article 12326. https://doi.org/10.3390/ijms252212326
- Korotkova, M., & Jakobsson, P. J. (2014). Characterization of microsomal prostaglandin E synthase 1 inhibitors. *Basic & Clinical*

- *Pharmacology & Toxicology*, *114*(1), 64-69. https://doi.org/10.1111/bcpt.12162
- Lin, P. C., Lin, Y. J., Lee, C. T., Liu, H. S., & Lee, J. C. (2013). Cyclooxygenase-2 expression in the tumor environment is associated with poor prognosis in colorectal cancer patients. *Oncology Letters*, 6(3), 733-739. https://doi.org/10.3892/ol.2013.1426
- Masae, M., Nasawat, A., Sangsakul, N., Mansoh, N., & Ruanglertboon, W. (2025). Exploring the clinical characteristics and survival outcomes in colorectal cancer patients in Hatyai hospital: A retrospective cohort study. *Journal of Current Science and Technology*, 15(3), Article 123.
 - https://doi.org/10.59796/jcst.V15N3.2025.123
- Muthukumaradoss, K., Babu, S., Vellapandian, C., Vellapandian, C., & Palaniappan, S. (2022). The biology and chemistry of microsomal prostaglandin E synthase (mPGES)-I inhibitors for cancer biomedicine. In *Systems Biomedicine Approaches in Cancer Research*. Singapore: Springer Nature Singapore. https://doi.org/10.1007/978-981-19-1953-4_4
- Mohammed, A., & Shoemaker, R. H. (2022).

 Targeting the leukotriene pathway for colon cancer interception. *Cancer Prevention Research*, *15*(10), 637-640.

 https://doi.org/10.1158/1940-6207.CAPR-22-0331
- Murakami, M., & Kudo, I. (2006). Prostaglandin E synthase: A novel drug target for inflammation and cancer. *Current Pharmaceutical Design*, *12*(8), 943-954. https://doi.org/10.2174/138161206776055912
- Mowat, A. N., & Al-Abady, Z. N. (2025). Study the Impact of NADPH oxidase1, TNF-alpha and Cyclooxygenase-2 On Colon Cancer Patients. *Asian Pacific Journal of Cancer Prevention: APJCP*, 26(3), 877-887.
 - https://doi.org/10.31557/APJCP.2025.26.3.877
- Myung, S. J., Rerko, R. M., Yan, M., Platzer, P., Guda, K., Lawrence, E.,... & Markowitz, S. D. (2006). 15-Hydroxyprostaglandin dehydrogenase is an in vivo suppressor of colon tumorigenesis. *Proceedings of the National Academy of Sciences of the United States of America*, 103(32), 12098–12102. https://doi.org/10.1073/pnas.0603235103
- Nanda, N., & Dhawan, D. K. (2021). Role of cyclooxygenase-2 in colorectal cancer. *Frontiers in Bioscience-landmark*, 26(4), 706-716.

- https://www.imrpress.com/journal/fbl/26/4/10.2 741/4914
- Negi, R. R., Rana, S. V., Gupta, V., Gupta, R., Chadha, V. D., Prasad, K. K., & Dhawan, D. K. (2019). Over-expression of cyclooxygenase-2 in colorectal cancer patients. *Asian Pacific Journal of Cancer Prevention: APJCP*, 20(6), 1675-1681. https://doi.org/10.31557/APJCP.2019.20.6.1675
- Purnama, A., Lukman, K., Rudiman, R., Prasetyo, D., Fuadah, Y., Nugraha, P., & Candrawinata, V. S. (2023). The prognostic value of COX-2 in predicting metastasis of patients with colorectal cancer: A systematic review and meta analysis. *Heliyon*, *9*(10), Article e21051. https://doi.org/10.1016/j.heliyon.2023.e21051
- Radhi, A. A., Muslim, O. T., & Abdlmaged, M. A. (2018). Epidemiological distribution of colorectal cancer in AL-Diwaniyah province, Iraq: An observational study. *Journal of Pharmaceutical Sciences and Research*, 10(7), 1758-1760.
- Rahman, M., Selvarajan, K., Hasan, M. R., Chan, A. P., Jin, C., Kim, J., ... & Tai, I. T. (2012). Inhibition of COX-2 in colon cancer modulates tumor growth and MDR-1 expression to enhance tumor regression in therapyrefractory cancers in vivo. *Neoplasia*, *14*(7), 624-633. https://doi.org/10.1593/neo.12486
- Ricciotti, E., & FitzGerald, G. A. (2011).

 Prostaglandins and inflammation. *Arteriosclerosis, Thrombosis, and Vascular Biology*, *31*(5), 986-1000.

 https://doi.org/10.1161/ATVBAHA.110.207449
- Roelofs, H. M., Te Morsche, R. H., van Heumen, B. W., Nagengast, F. M., & Peters, W. H. (2014). Over-expression of COX-2 mRNA in colorectal cancer. *BMC Gastroenterology*, *14*(1), Article 9005. https://doi.org/10.1186/1471-230X-14-1
- Sabah, Y. H., & Hassan, F. (2024). Comparison of serum vitamins C, D, and E in Iraqi colorectal cancer patients with and without non-alcoholic fatty liver disease to healthy individuals. *Journal of Current Science and Technology*, 14(3), Article 56. https://doi.org/10.59796/jcst.V14N3.2024.56
- Sasaki, Y., Nakatani, Y., & Hara, S. (2015). Role of microsomal prostaglandin E synthase-1 (mPGES-1)-derived prostaglandin E2 in colon carcinogenesis. *Prostaglandins & Other Lipid Mediators*, 121, 42-45.

- https://doi.org/10.1016/j.prostaglandins.2015. 06.006
- Seo, T., Tatsuguchi, A., Shinji, S., Yonezawa, M., Mitsui, K., Tanaka, S., ... & Sakamoto, C. (2009). Microsomal prostaglandin E synthase protein levels correlate with prognosis in colorectal cancer patients. *Virchows Archiv*, 454(6), 667-676. https://doi.org/10.1007/s00428-009-0777-z
- Serrano-López, J., & Martín-Antonio, B. (2021). Inflammaging, an imbalanced immune response that needs to be restored for cancer prevention and treatment in the elderly. *Cells*, *10*(10), Article 2562. https://doi.org/10.3390/cells10102562
- Shalaby, S. M., Shawky, S. A., Ashour, H., & Sarhan, W. (2025). The interplay between COX-2, chemotherapeutic drugs, and chemoresistance in colon cancer. *Scientific Reports*, *15*(1), Article 15837. https://doi.org/10.1038/s41598-025-98451-9
- Shaukat, A., Kahi, C. J., Burke, C. A., Rabeneck, L., Sauer, B. G., & Rex, D. K. (2021). ACG clinical guidelines: Colorectal cancer screening 2021. *The American Journal of Gastroenterology*, 116(3), 458-479. https://doi.org/10.14309/ajg.00000000000001122
- Sheng, J., Sun, H., Yu, F. B., Li, B., Zhang, Y., & Zhu, Y. T. (2020). The role of cyclooxygenase-2 in colorectal cancer. *International Journal of Medical Sciences*, 17(8), Article 1095. https://doi.org/10.7150/ijms.44439
- Siegel, R. L., Miller, K. D., Goding Sauer, A., Fedewa, S. A., Butterly, L. F., Anderson, J. C., ... & Jemal, A. (2020). Colorectal cancer statistics, 2020. *CA: A Cancer Journal for Clinicians*, 70(3), 145-164. https://doi.org/10.3322/caac.21601
- Siegel, R. L., Wagle, N. S., Cercek, A., Smith, R. A., & Jemal, A. (2023). Colorectal cancer statistics, 2023. *CA: A Cancer Journal for*

- *Clinicians*, 73(3), 233-254. https://doi.org/10.3322/caac.21772
- Sophonnithiprasert, T., Konjanthet, S., Narinnork, N., Prompat, N., Benjakul, S., Saetang, J., ... & Rattanaburee, T. (2025). B-AP15 inhibited colon cancer cell proliferation by decreasing CDK6, Cyclin A, Cyclin E, c-Myc, and VEGF gene expression. *Naunyn-Schmiedeberg's Archives of Pharmacology*, 1-16. https://doi.org/10.1007/s00210-025-03940-3
- Stamatakis, K., Jimenez-Martinez, M., Jimenez-Segovia, A., Chico-Calero, I., Conde, E., Galán-Martínez, J., ... & Fresno, M. (2015). Prostaglandins induce early growth response 1 transcription factor mediated microsomal prostaglandin E2 synthase up-regulation for colorectal cancer progression. *Oncotarget*, 6(37), 39941–39959. https://doi.org/10.18632/oncotarget.5402
- Szweda, M., Rychlik, A., Babińska, I., & Pomianowski, A. (2019). The significance of cyclooxygenase-2 in oncogenesis:
 Cyclooxygenase-2 as a biomarker with diagnostic, therapeutic, prognostic, and predictive relevance in small animal oncology.

 Journal of Veterinary Research, 63(2), 215–224. https://doi.org/10.2478/jvetres-2019-0030
- Yang, W., Luo, Y., Hu, S., Li, Y., & Liu, Q. (2018). Value of combined detection of serum carcino-embryonic antigen, carbohydrate antigen 19-9 and cyclooxygenase-2 in the diagnosis of colorectal cancer. *Oncology Letters*, 16(2), 1551-1556. https://doi.org/10.3892/ol.2018.8792
- Yarla, N. S., Madka, V., Pathuri, G., & Rao, C. V. (2020). Molecular targets in precision chemoprevention of colorectal cancer: An update from pre-clinical to clinical trials. *International Journal of Molecular Sciences*, 21(24), Article 9609. https://doi.org/10.3390/ijms21249609